

Multiplex molecular infection diagnostic – Complete typing of human papilloma viruses

EUROIMMUN

Medizinische
Labor Diagnostika
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Topics of the talk

- Overview of current non-cultural methods for pathogen detection
- Development of a PCR based multiparameter assay



Non-cultural methods for pathogen diagnostics are fast and standardized

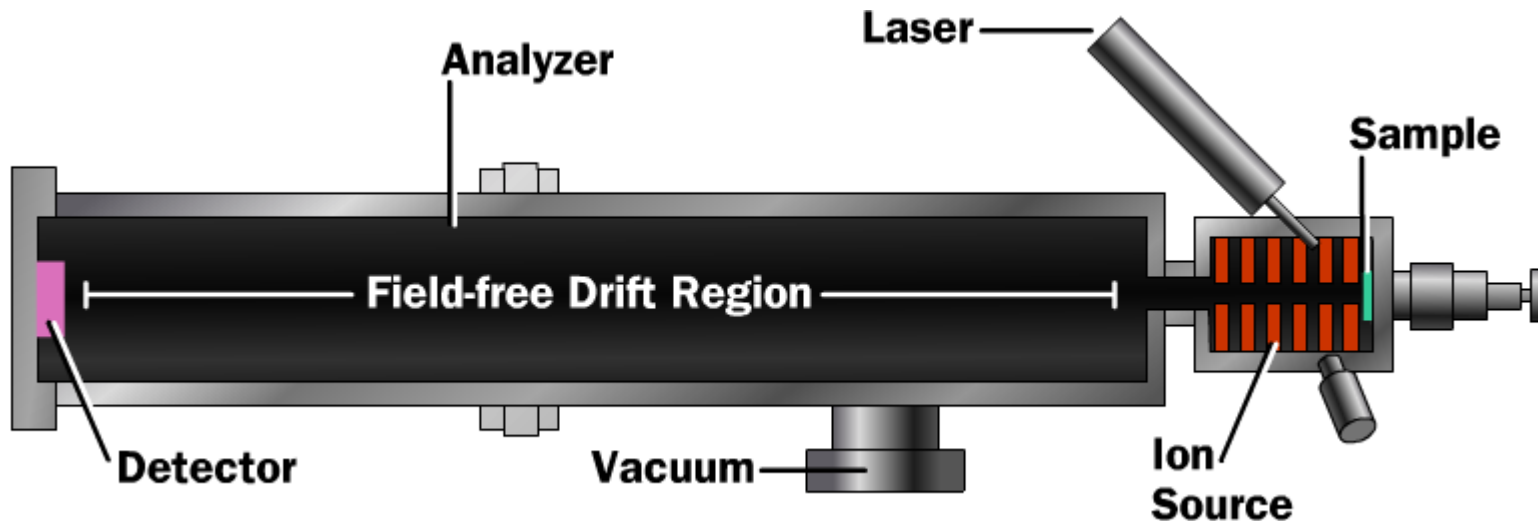
Short overview of current non-cultural methods for pathogen detection

- Maldi-Tof
- PCR-based Methods
 - Real-Time PCR
 - Sequencing
 - Microarray technology



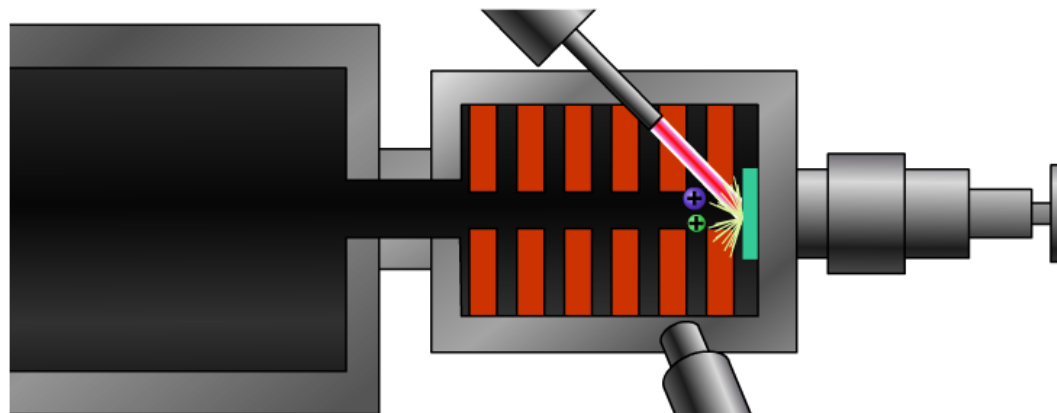
Maldi-Tof allows the analysis of biomolecules when ionized

- Maldi-Tof = Matrix-assisted laser desorption/ionization time-of-flight mass spectrometer



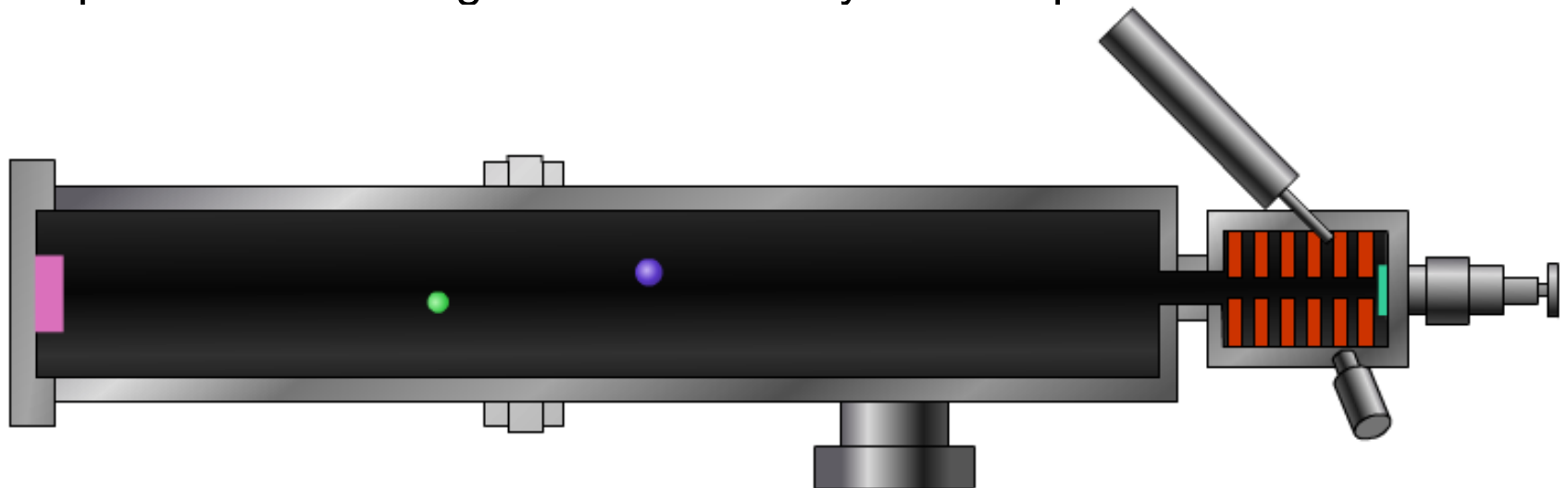
Biomolecules are separated based on their difference in mass

- The analyte will be co-crystallized with a matrix compound, usually a UV-absorbing organic acid
- The matrix absorbs UV-light and converts it to heat energy
- The matrix becomes ionized with a single positive charge, this positive charge is transferred to native sample proteins through their random collision in the gas phase
- Because all native sample proteins sample have an identical, single positive charge, they are separated based on their difference in mass



The sample will be identified in comparison to data of known organisms

- Heavier ions will travel through the mass analyzer at a slower velocity, compared to lighter ions
- An ion detector measures the time to impact
- Based on standards of known mass, the time to impact for each unknown analyte is converted into a mass-to-charge ratio
- Pattern of proteins will be compared with a database of Maldi-Tof spectra of known organisms to identify the sample

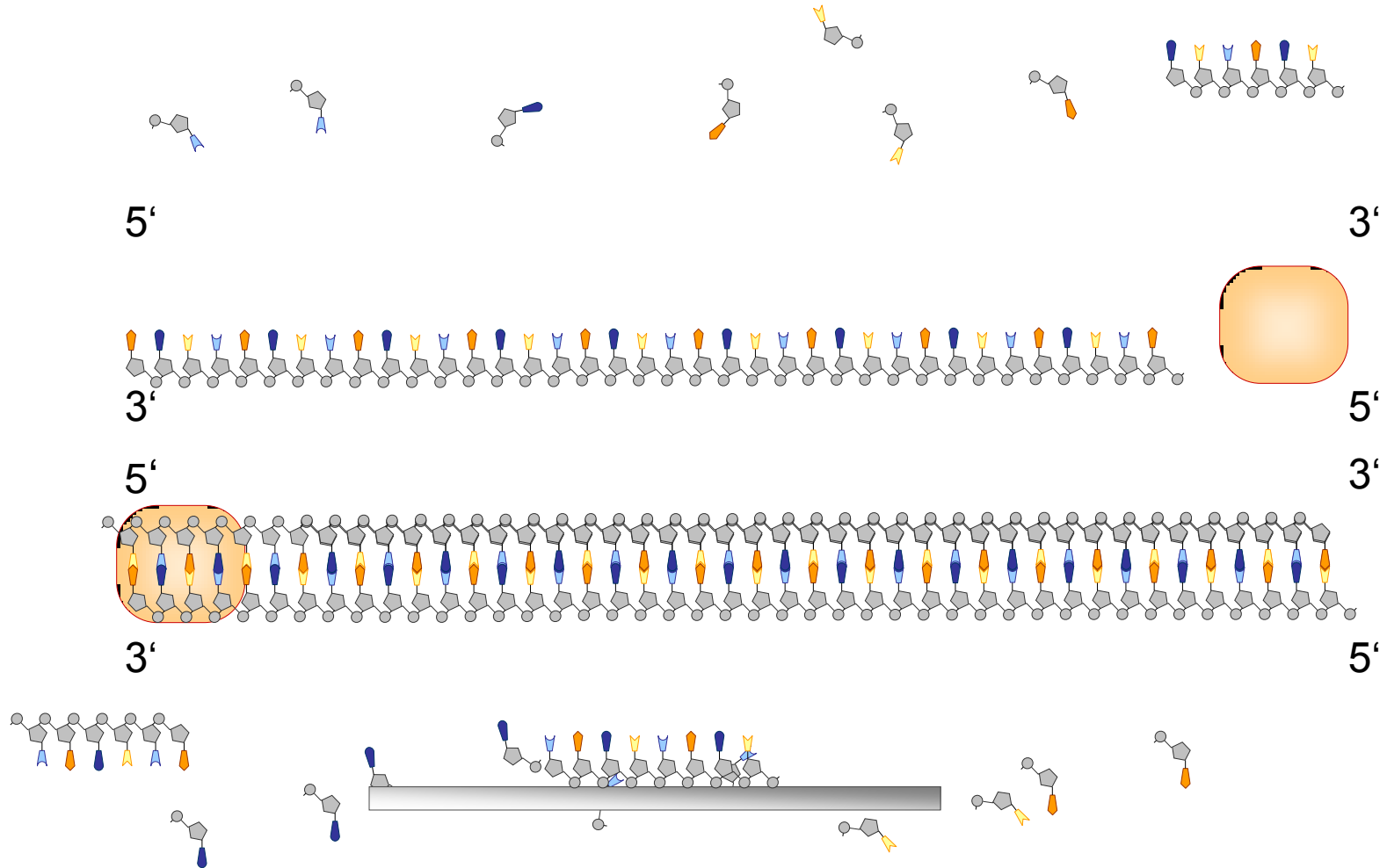


Advantages and disadvantages of Maldi-Tof analysis

- Advantages
 - Easy
 - Price/analysis is cheap
- Disadvantages
 - High investment costs
 - No resistance gene detection
 - Need of pure culture
 - Only protein patterns which are stored in a database may give an answer



PCR-based methods – Principles of Polymerase chain reaction



The selection of PCR-components is not unimportant

- There are various polymerase enzymes on the market
 - Speed
 - Quality of amplification
 - Stability
 - ...
- The compounds shouldn't be contaminated with the DNA which have to be amplified



PCR - one basic technique – different evaluation methods

Sequencing

Real-Time PCR

- Two common methods
 - (1) non-specific fluorescent dyes that intercalate with any double-stranded DNA while PCR
 - (2) sequence-specific labelled DNA probes

Microarray

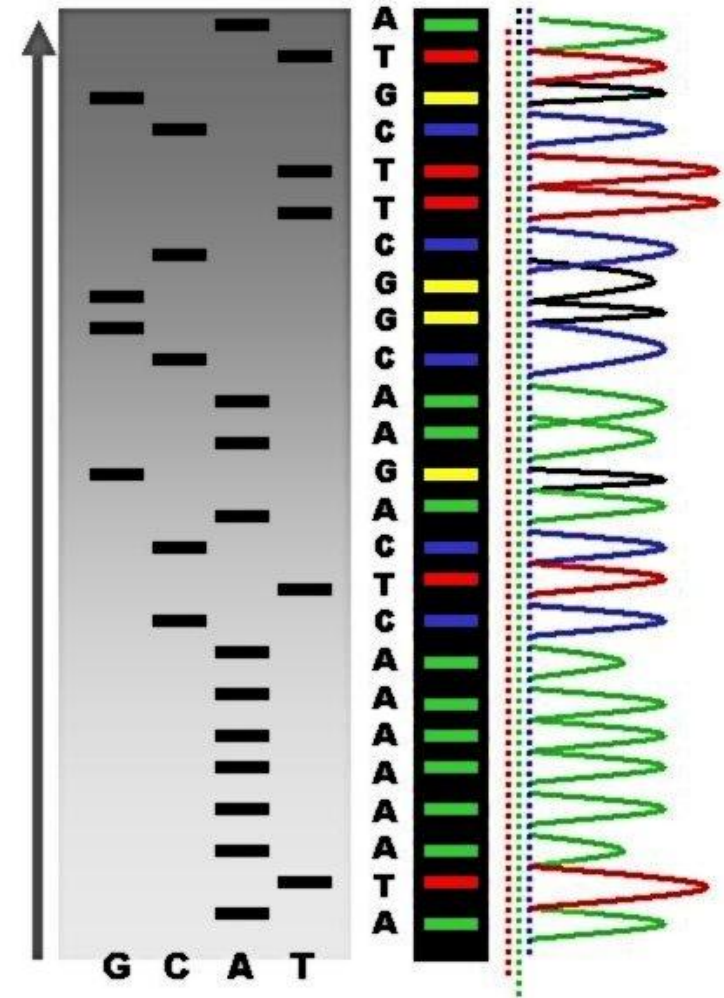
- Amplification of the DNA by PCR
- Analysis via specific probe binding on an array



Sequencing is the process of determining the nucleotide order of a DNA fragment

Sanger Sequencing

- Chain termination method
- Uses sequence-specific termination of a DNA synthesis during the PCR
- The knowledge about the terminating modified nucleotide shows the sequence of the DNA fragment



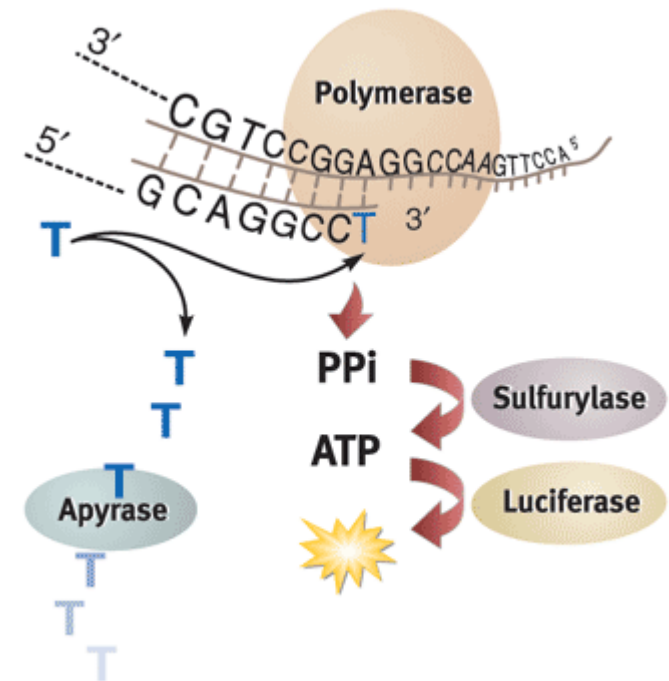
New sequencing technologies are gaining an increasing share of the sequencing market

Next-generation sequencing methods

Whole genomes can be sequenced in a single run with several times coverage

Example: Pyrosequencing

- DNA is annealed to beads and amplified via emulsion-based clonal amplification
- Free nucleotides are washed over the DNA
- ATP is generated when nucleotides join with their complementary base pairs
- Enzymes produce light in the presence of ATP
- The signal strength is proportional to the number of nucleotides, incorporated in a single nucleotide flow



Advantages and disadvantages of sequencing methods

- Advantages
 - All information in one step
 - Also new mutations are detected
- Disadvantages
 - High investment costs
 - Expensive
 - Need of pure culture
 - Depending on the method high amount of data and more information then requested



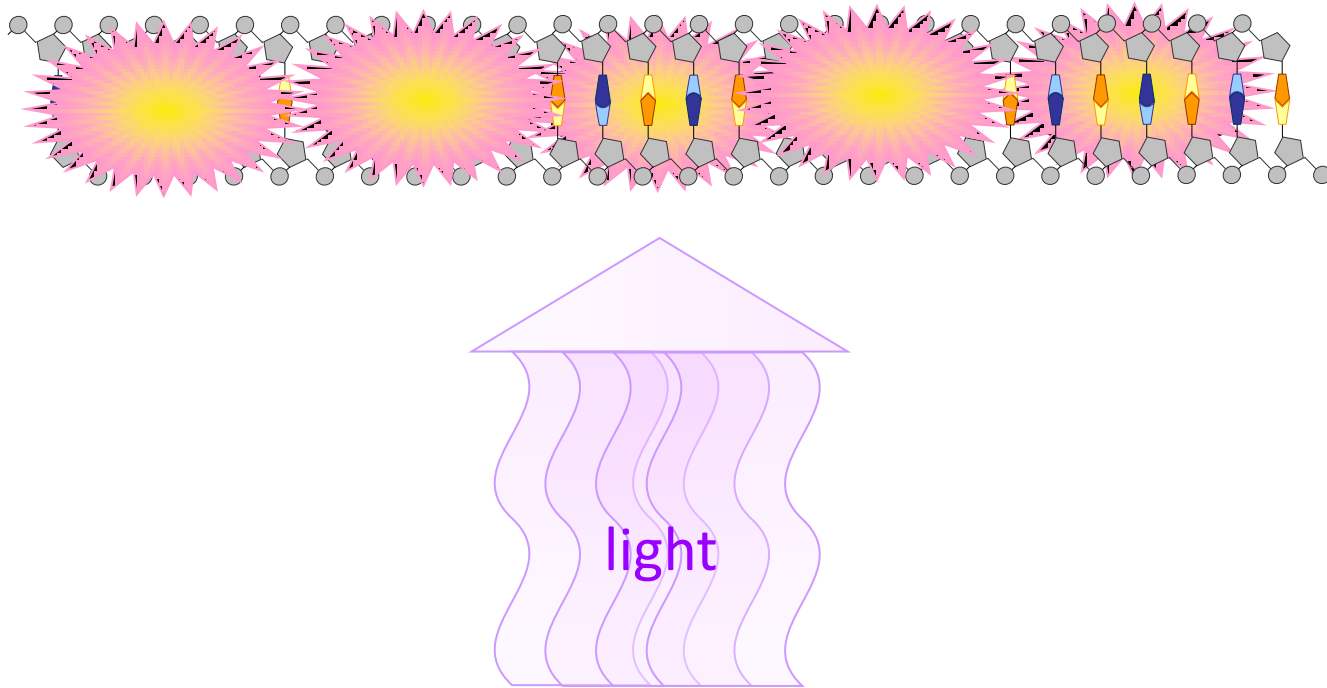
Real-Time PCR monitors the amplification of a targeted DNA molecule during the PCR

- There are two ways for the direct detection of PCR products while RT-PCR
 - non-specific fluorescent dyes that intercalate with any double-stranded DNA while PCR
 - sequence-specific labelled DNA probes



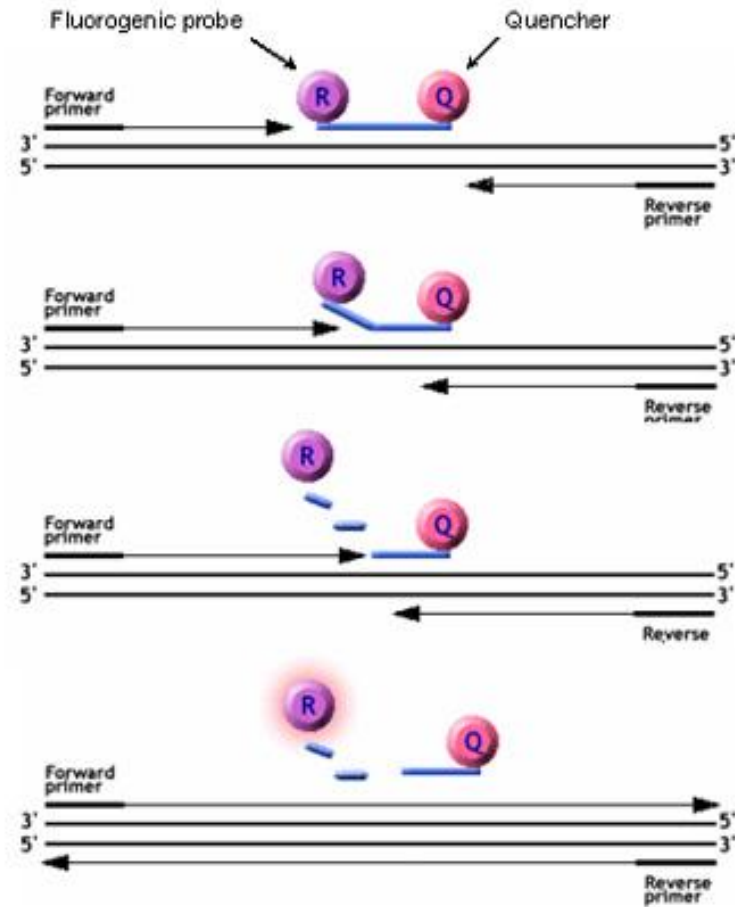
Easy but no additional specificity – intercalating dyes

- Intercalating dyes like SybrGreen give a fluorescent signal by activation with light



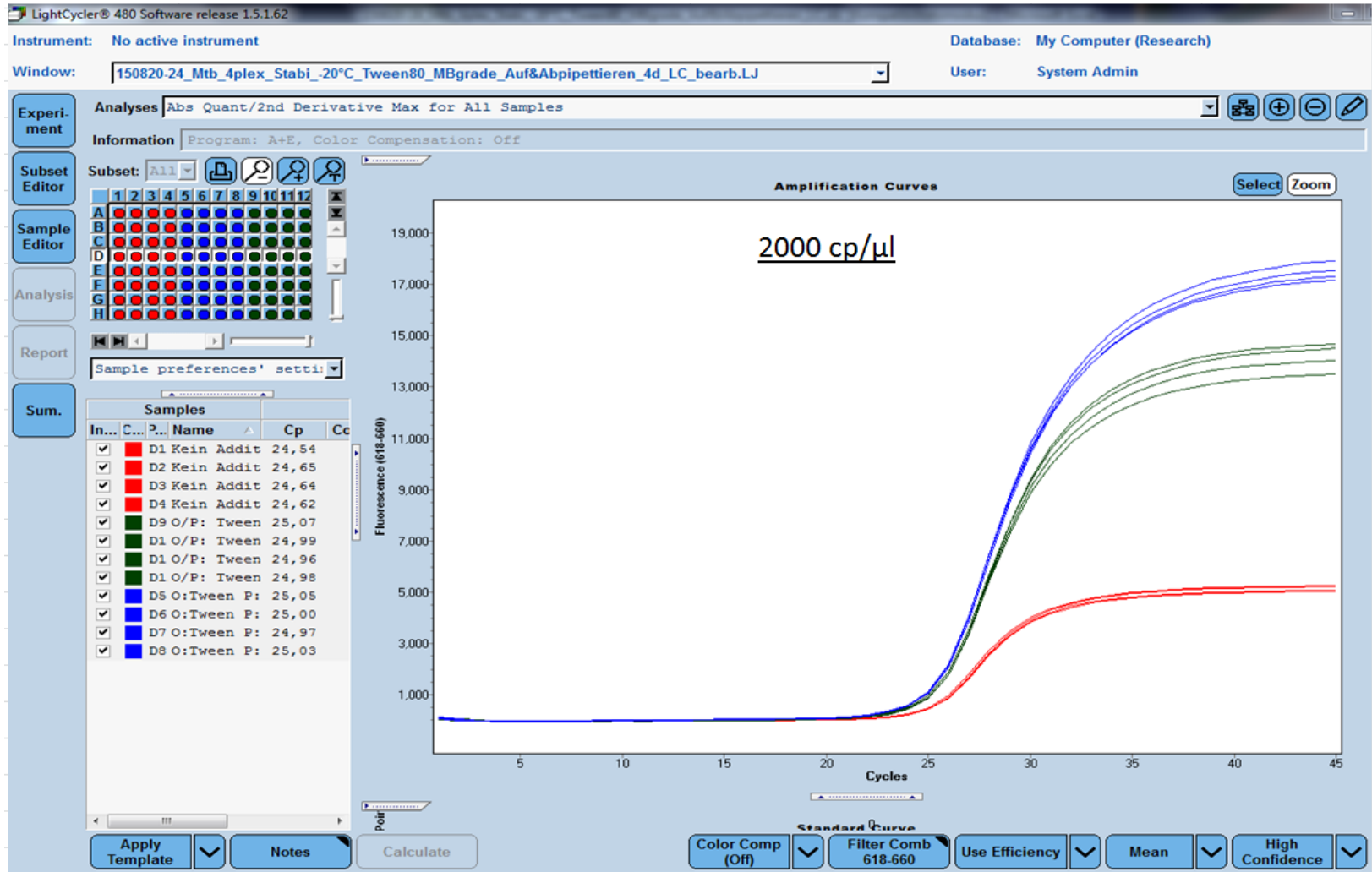
Real-Time PCR with dye-labeled probes – two step specificity possible

- Fluorescence of the reporter dye is prevented by the quencher
- Probe has to bind to its specific, complementary sequence
- As the DNA polymerase moves along the template, the probe is cleaved (broken) between the reporter and quencher
- This allows the reporter dye to emit fluorescence as it is no longer suppressed by the quencher dye
- Reporter fluorescence increases during each PCR cycle and is proportional to the amount of PCR product.



Hawrami, K and Brewer, J (1997) *Journal of Medical Virology*, 53 pp60-63

The typical outcome of a RT-PCR is an amplification curve



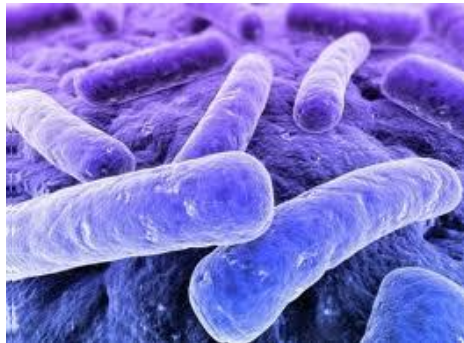
Advantages and disadvantages of RT-PCR

- Advantages
 - Amplifying and detection in one step
- Disadvantages
 - Quantification of DNA/RNA copies possible
 - Number of test parameters limited



Challenges of the molecular infection diagnostics

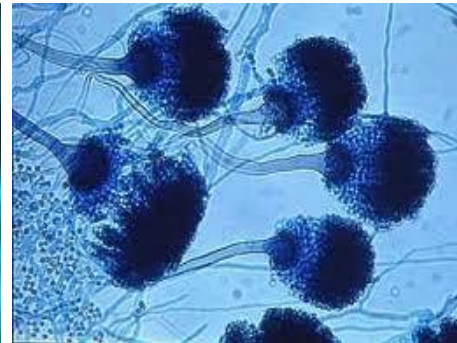
- Direct analysis of pathogens and resistant gens by the detection of the DNA with μ Array technology



Bacteria



Virus



Fungus



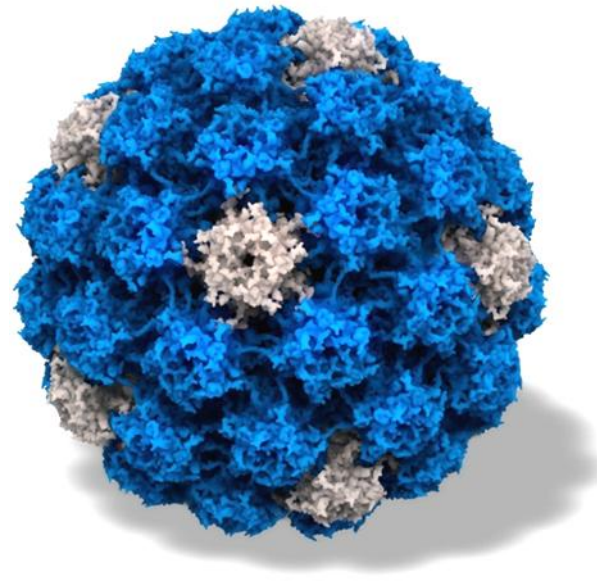
Parasites



Microarrays in the CE/IVD diagnostics - Development of an HPV Array

„There is a sufficient evidence that testing for human Papillomavirus infection as the primary screening modality can reduce cervical cancer incidence and mortality rates”

[International Agency for Research on Cancer – IARC, Handbooks of Cancer Prevention Cervical Cancer Prevention, Volume 10, IARC Press 2005]



HPV are the most common sexual transmitted virus

- HPV are the most common pathogens of STD
- An HPV infection very often occur already during the first sexual contacts
- The HPV prevalence vary in the population depending on the age, the social stratum, the culture group an the associated sexual behavior between 3% und 50%
(Munoz et al., 1996; van den Brule et al., 1991; Schneider et al., 2000)



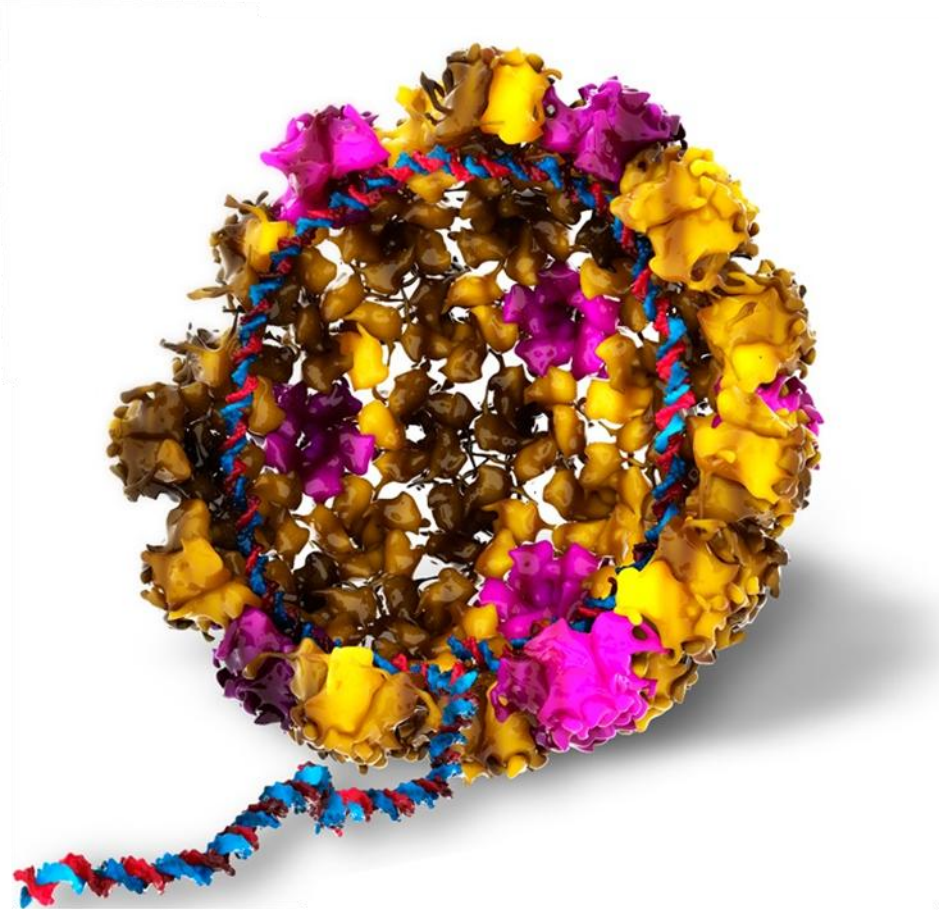
HPV - some backgrounds

- HPV are double stranded DNA virus
- The genome size is approximately 8000 bp
- ~210 humanpathogene HPV-subtypes are described
(http://pave.niaid.nih.gov/#explore/reference_genomes/human_genomes)
- HPV infection is limited to the basal cells squamous epithelium of skin and mucosa
- The viral replication is only possible in fully differentiated squamous epithelium
- After the infection the viral DNA will be replicated extrachromosomal in the cell nucleus of the host cell



The oncogenic potential of the HPV is mediated by two genes

- The double stranded circular HPV DNA is organized in 9 genes
 - Variable number of early genes: e.g. **E6**, **E7**, E1, E2, E3, E4, E5
 - 2 late genes: L1, L2
- High-risk HPV and low-risk HPV have different abilities to influence the cell cycle by the inhibition or deactivation of cell cycles regulating proteins because of their variants of the E6 and E7 protein



The HPV genome is subdivided in a LCR, early and late gens

- Non-coding-long-control-region (LCR)
 - Promotor region for the control of the gen expression of the HPV gens (early-gens and late-gens)
- Early-gens
 - Not very conservative
 - Regulatory gen products
 - Necessary for the process of the malign degeneration of the host cell
- Late-gens
 - Conservative
 - Coding for the viral capsid proteins L1 and L2
- HPV genes will be transcribed as polycistronic RNA with overlapping open-reading-frames (ORF)



30 anogenital HPV are known

- 30 HPV are known which infect exclusively the skin and the mucosa of the anogenital region
- Anogenital HPV are sub-classified in two groups

(1) Low-risk-HPV subtypes¹

- HPV 6, 11, 40, 42, 43, 44, 54, 61, 70, 72, 81 und 89 (CP6108)

(2) High-risk-HPV subtypes^{1,2}

- Identified in 99,7% of all cervical cancer tumors
- These days most of cervical carcinoma (~ 70%) provoked by high-risk HPV subtypes 16 and 18
- HPV 16, 18, 26, 31, 33, 35, 39, 45, 51, 52, 53, 56, 58, 59, 66, 68, 73 und 82

¹Munoz,N., Bosch,F.X., de,S.S., Herrero,R., Castellsague,X., Shah,K.V., Snijders,P.J., and Meijer,C.J. (2003). Epidemiologic classification of human papillomavirus types associated with cervical cancer. *N Engl J Med* 348, 518-527

Official classified as carcinogenic by the WHO, Cogliano,V., Baan,R., Straif,K., Grosse,Y., Secretan,B., and El,G.F. (2005). Carcinogenicity of human papillomaviruses. *Lancet Oncol.* 6, 204



HPV are able to induce cellular transformations

- Some HPV subtypes can induce (malign) cellular transformations
 - Cervical cancer
 - Vagina carcinoma
 - Penis carcinoma
 - Anal carcinoma
 - Carcinoma of the oral mucosa



Basis of PCR reactions – the primer and probe design

- Some key-proteins (and their nucleic acid sequences) stayed conserved thru evolution and differ in only few bp
 - For example: ribosomal subunits
 - With primers/probes for conserved DNA sequences from a bundle of organisms will be amplified
- Some proteins (and their nucleic acid sequences) are unique and only present in one organism
 - With primers/probes for unique regions DNA sequences from only one organism will be amplified

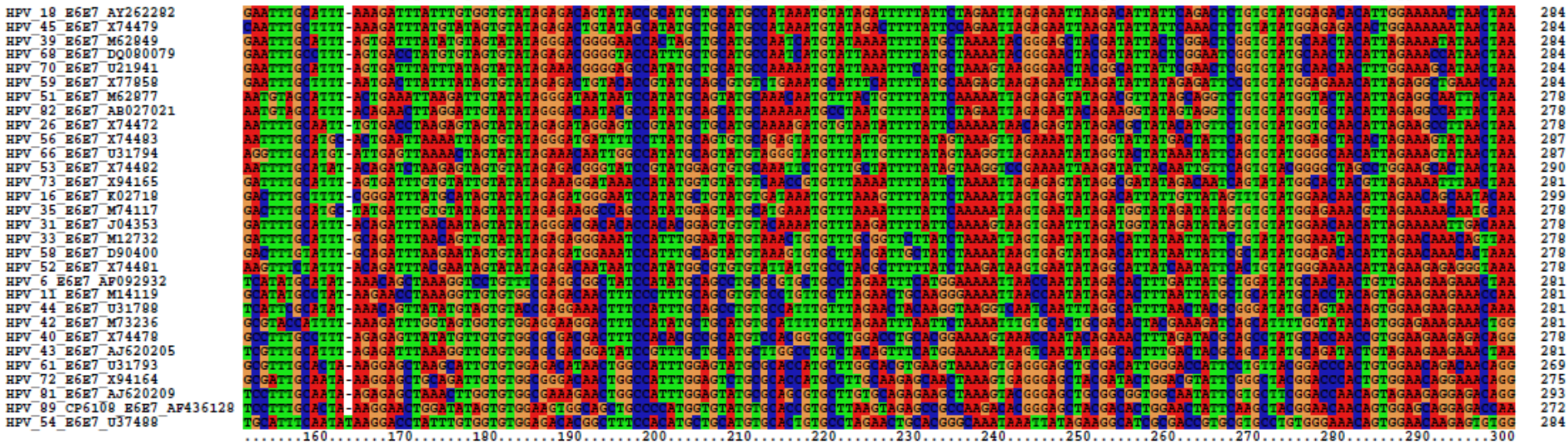


Depending on the pathogen panel different strategies for primers and probes are useful

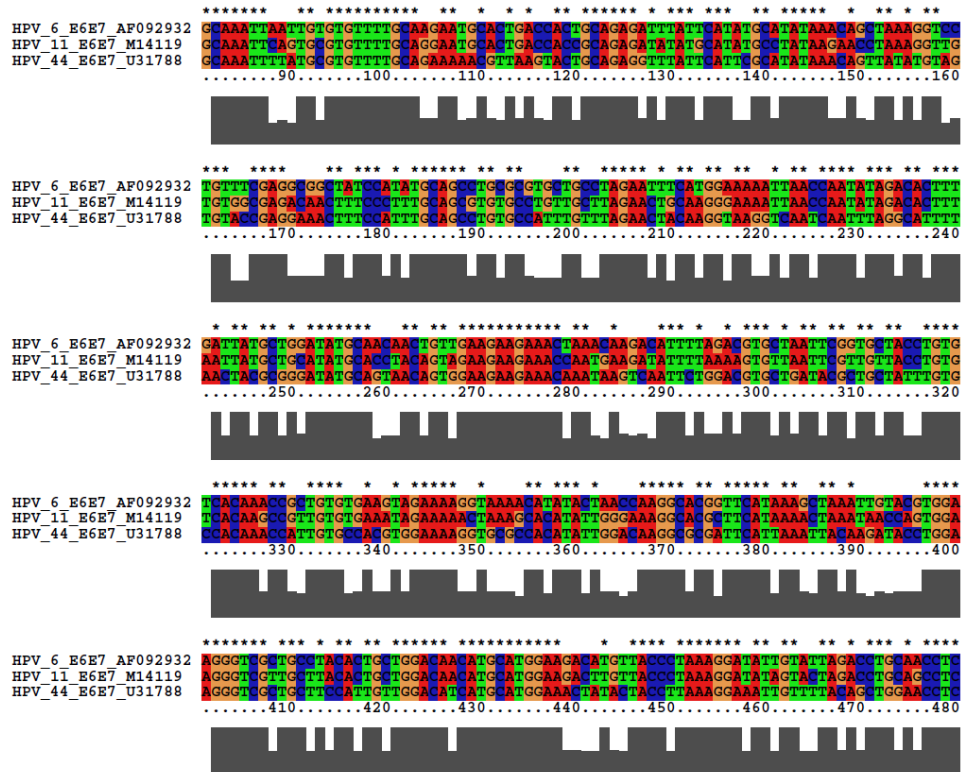
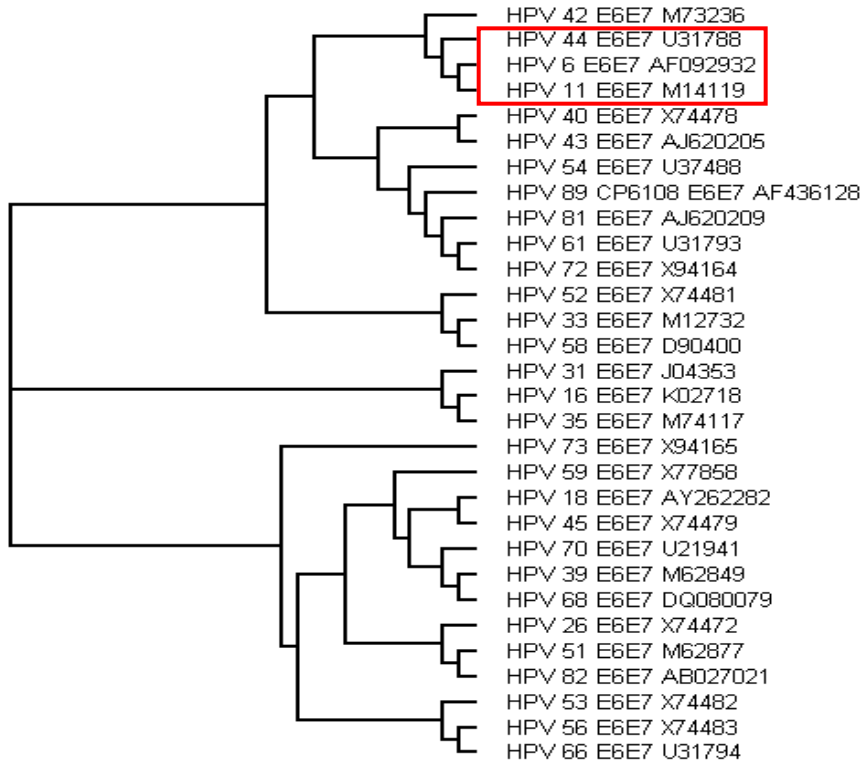
- Conservative primers and probes
 - Detection of all subtypes as a group with only a few primer/probe-systems
 - No subtyping possible
- Conservative primers, specific probes
 - Detection of all subtypes as a group with only a few primer
 - Specific probes give the chance for a limited subtyping
- Specific primers, specific probes
 - Detecting and full subtyping
 - Complex PCR-reactions with a lot of primers



In-silico work on HPV E6/E7 no conservative primer/probe-systems



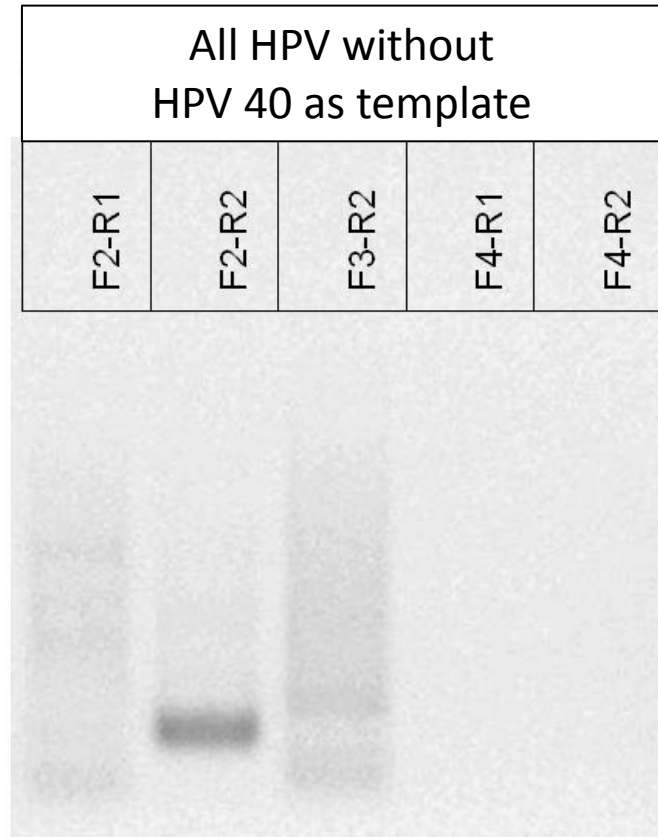
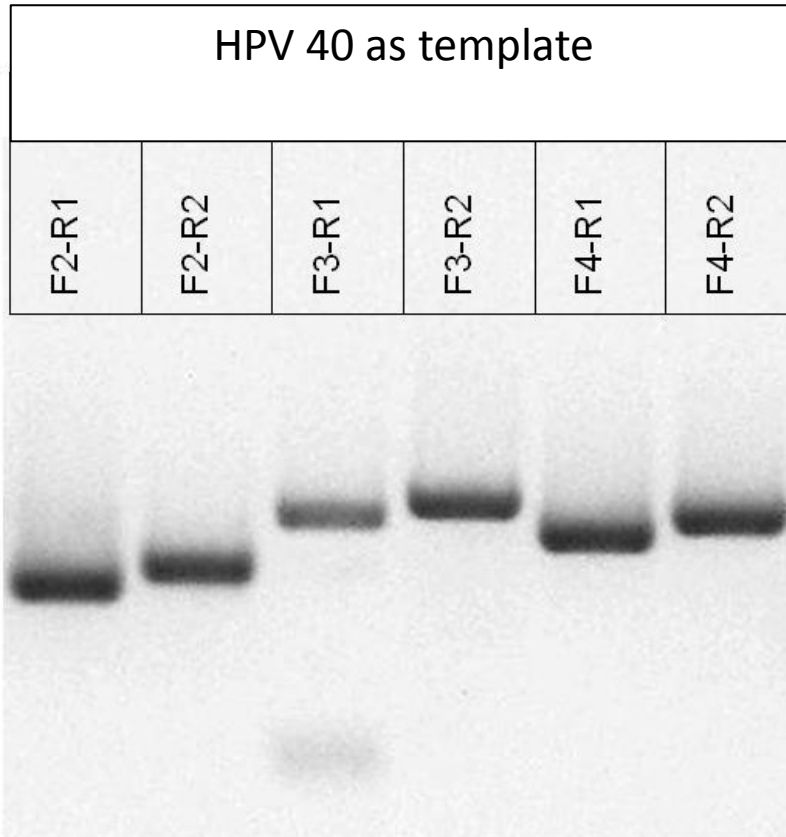
Smaler subgroups shows homologies - Selection of sequences for the development



Alignment der HPV Gene E6-E7

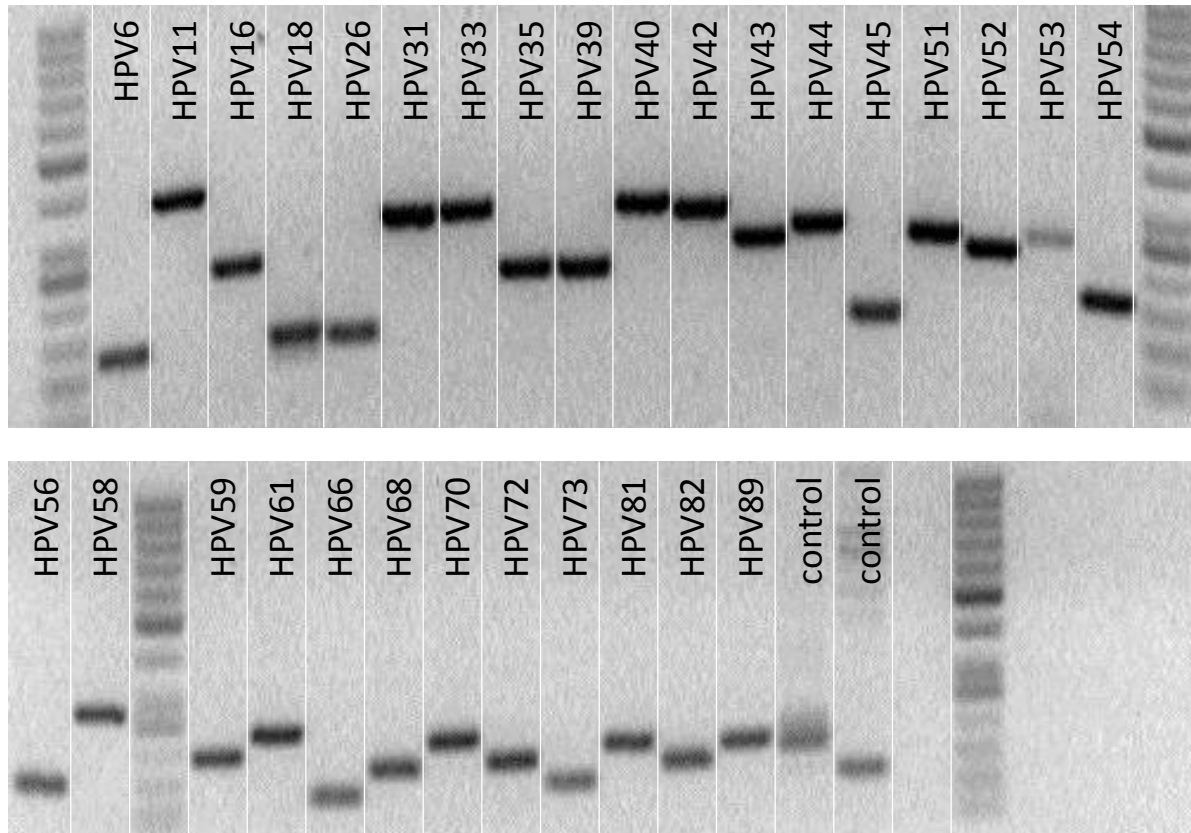


Example: development of a primer system for the HPV 40 detection

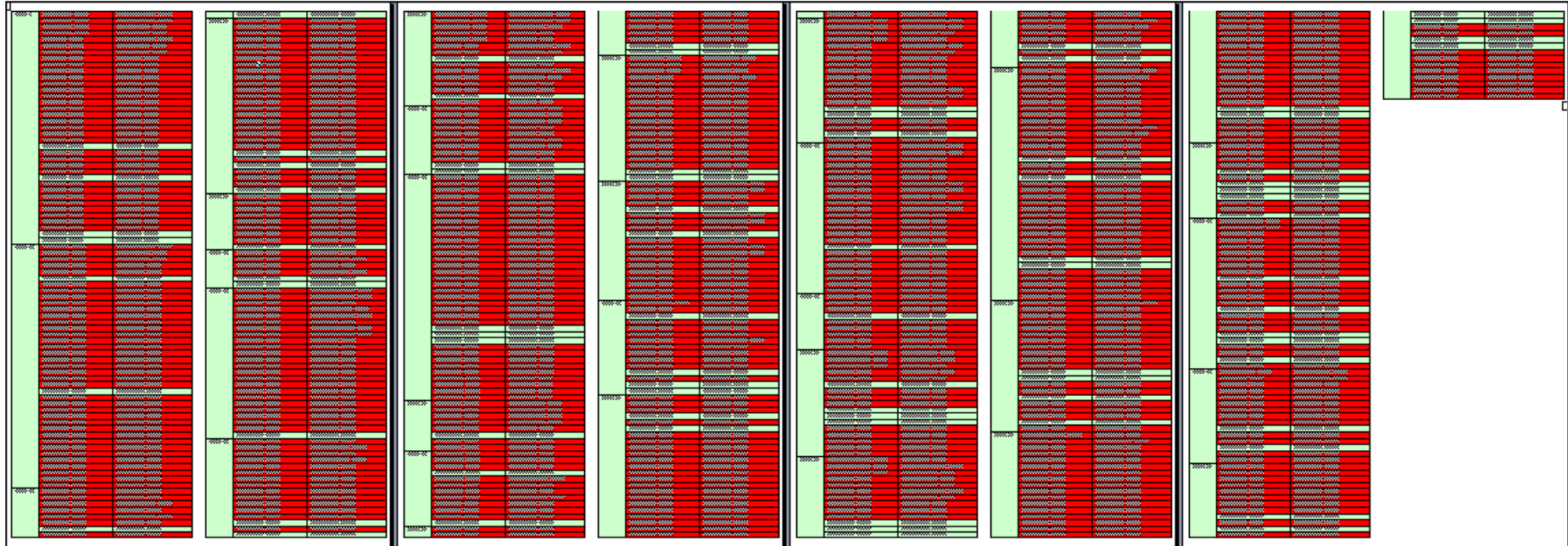


Optimizing of PCR-systems against cross reactions with single primer systems

Primer: single primer-system + Template: all 30 E6-E7 HPV-sequences + human genomic DNA



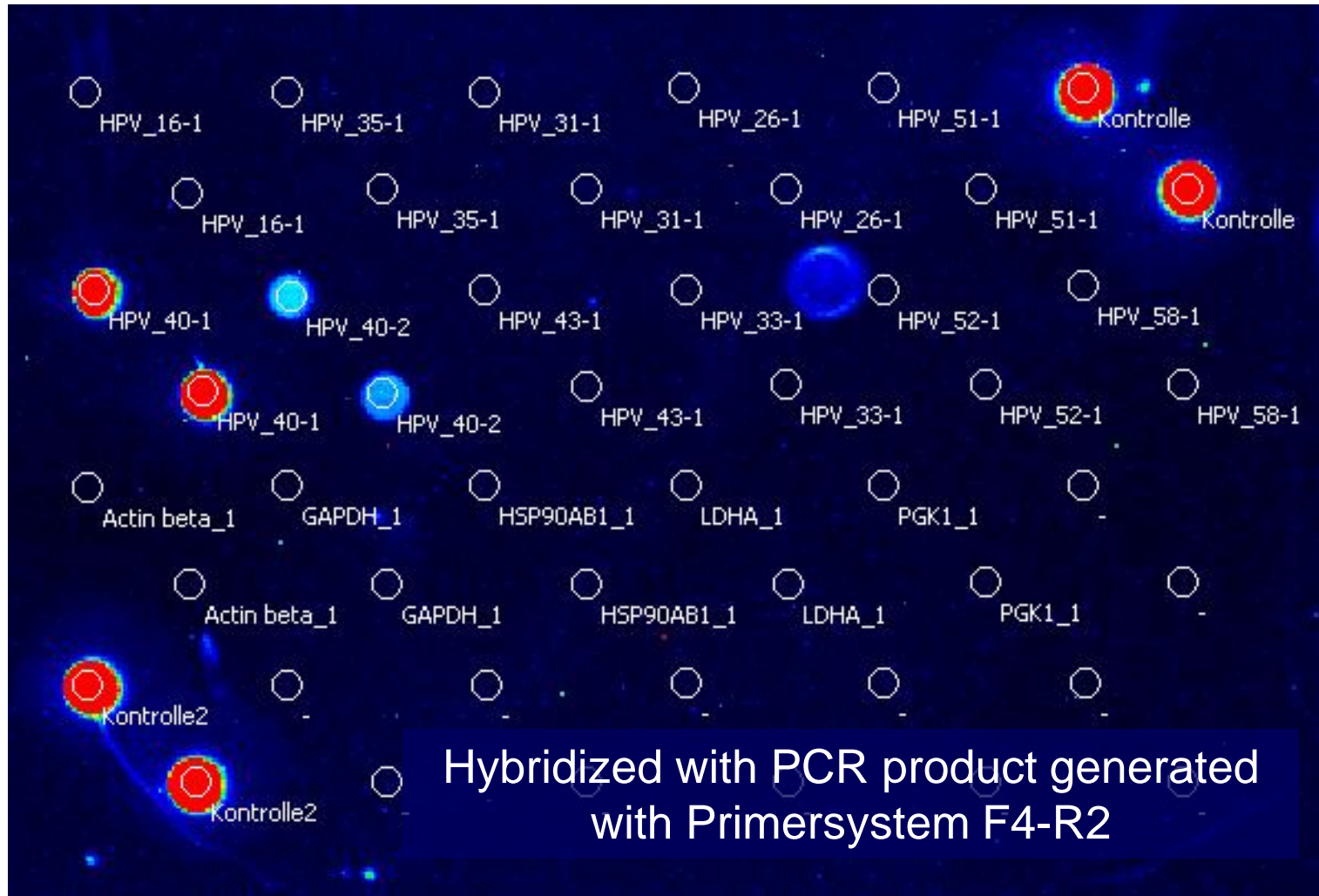
For a PCR with 31 primer pairs over 700 primer systems were developed and characterized



Over 700 tested primersystems



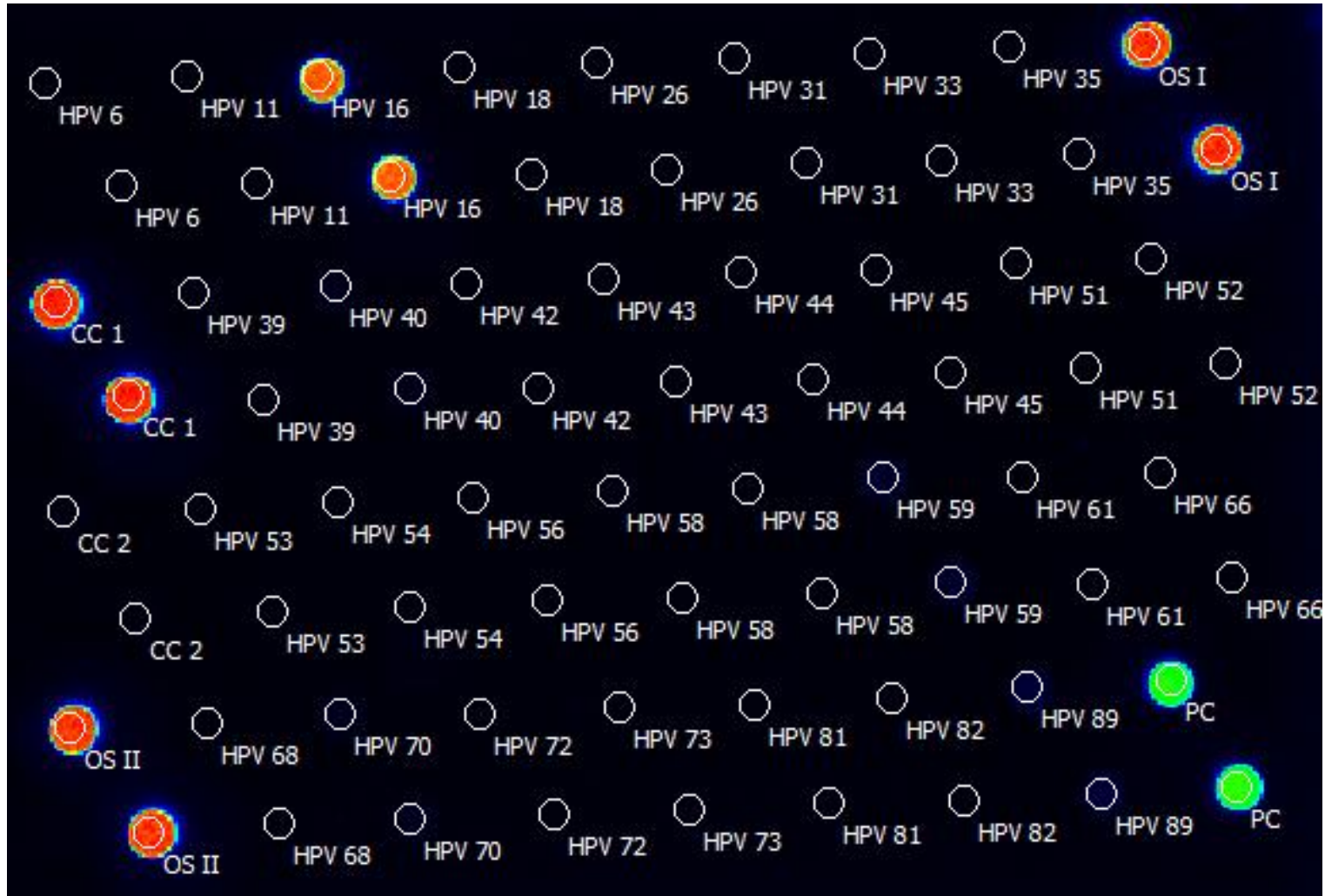
Example from the development: Selection of probes for HPV 40



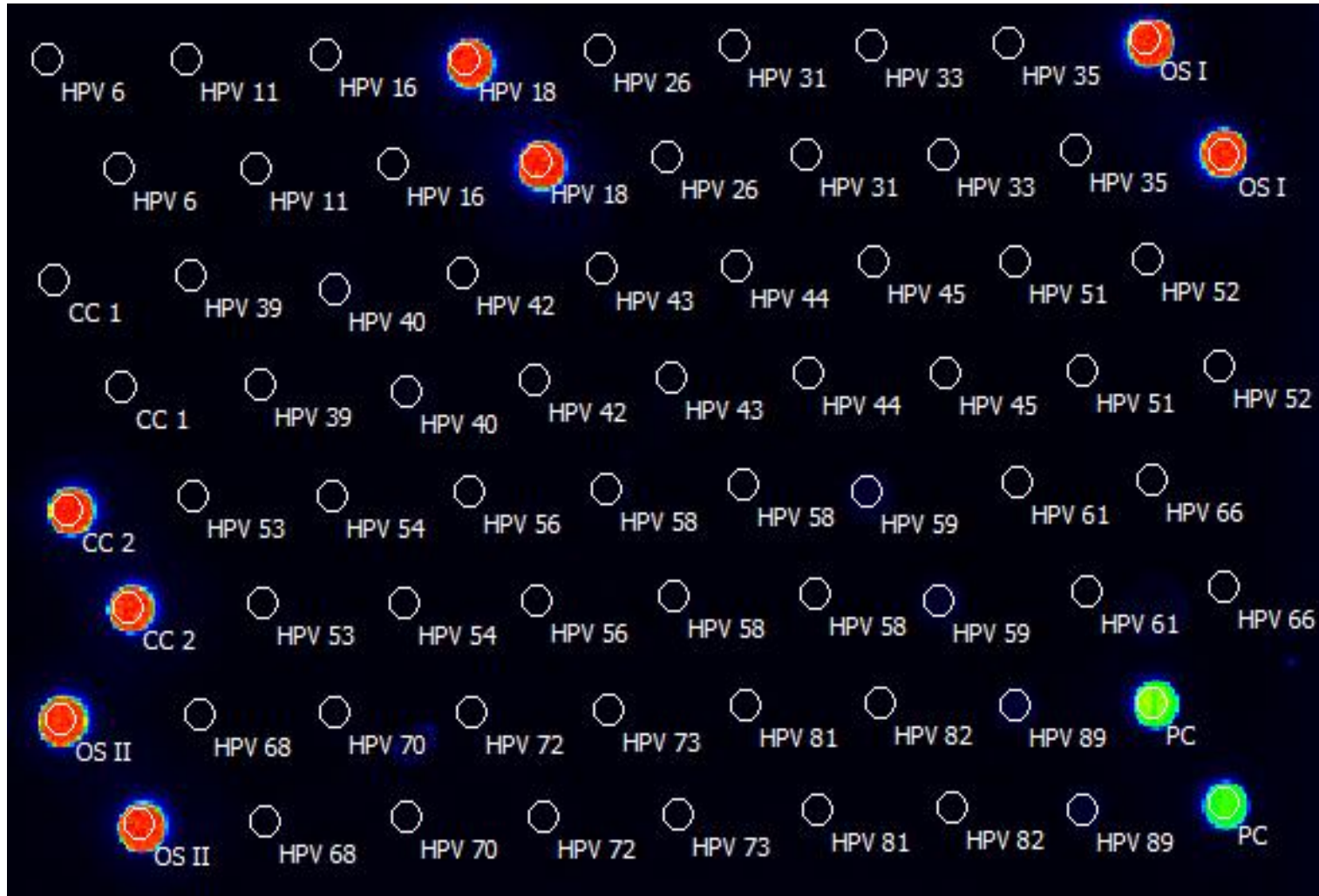
One primersystem for all HPV-subtypes + control = 62 primers - it works !!!

Template	HPV 6	HPV 11	HPV 16	HPV 18	HPV 26	HPV 31	HPV 33	HPV 35	HPV 39	HPV 42	HPV 43	HPV 44	HPV 45	HPV 51	HPV 52	HPV 53	HPV 54	HPV 56	HPV 58	HPV 59	HPV 61	HPV 66	HPV 68	HPV 72	HPV 73	HPV 81	HPV 82	HPV 89	PC	PC	DNA	NTC				
©	150	200	150	50	100	50	100	100	50	500	200	150	600	200	50	50	50	50	100	150	150	600	300	600	100	200	400	50	999	LoD	20x LoD	100 ng				
Patient-ID	HPV 6	HPV 11	HPV 16	HPV 18	HPV 26	HPV 31	HPV 33	HPV 35	HPV 39	HPV 42	HPV 43	HPV 44	HPV 45	HPV 51	HPV 52	HPV 53	HPV 54	HPV 56	HPV 58	HPV 59	HPV 61	HPV 66	HPV 68	HPV 72	HPV 73	HPV 81	HPV 82	HPV 89	PK	PK	PK	DNA	NTC			
	OT 1	Field	OT 1	Field	OT 1	Field	OT 2	Field	OT 2	Field	OT 3	Field	OT 3	Field	OT 4	Field	OT 4	Field	OT 5	Field	OT 5	Field	OT 6	Field	OT 6	Field	OT 7	Field	OT 7	Field	OT 7	Field	OT 7	Field		
HPV 6	38406	0	96	16	32	0	81	0	0	128	96	64	0	96	81	0	0	32	0	64	64	16	0	0	33	0	0	0	0	19644	59404	0	64			
HPV 6	43011	161	16	32	16	0	64	81	64	144	0	193	32	112	16	0	128	64	0	32	112	48	64	32	64	144	32	96	370	20880	59244	96	80			
HPV 11	0	2959	0	65	0	0	80	0	161	145	48	97	0	0	48	16	96	80	0	32	0	32	16	16	48	48	32	241	64	2568	12727	0	32			
HPV 11	0	3136	80	16	32	64	80	81	0	32	0	16	80	128	0	32	32	97	64	48	0	48	64	0	32	48	96	112	16	64	2648	11330	16	16		
HPV 16	80	0	2439	16	16	32	0	0	0	16	32	112	48	16	0	0	32	32	16	97	32	65	32	0	64	80	0	48	32	2167	33206	32	97			
HPV 16	80	0	2681	48	225	16	0	65	0	0	0	64	0	16	113	80	80	0	48	97	16	96	48	81	65	48	48	49	80	2343	35116	80	0			
HPV 18	321	337	288	41502	321	192	369	289	192	385	225	224	145	145	176	289	337	241	289	209	112	224	240	224	305	321	257	128	32	1476	10945	59468	193	17		
HPV 18	433	385	402	31537	369	209	369	273	225	305	273	224	225	128	193	337	273	273	289	193	225	225	289	337	304	257	145	209	321	10335	59404	209	0			
HPV 26	386	208	401	578	18504	289	353	241	304	337	514	176	160	112	80	305	337	257	193	241	129	273	272	289	353	257	192	224	289	8603	50105	177	49			
HPV 26	401	305	386	497	18152	273	273	225	289	256	337	160	225	209	257	321	192	256	241	193	225	273	273	321	273	305	177	273	177	305	7960	43412	177	32		
HPV 31	305	224	257	321	225	5103	257	257	144	257	321	305	128	161	305	417	257	369	257	241	225	209	272	386	273	321	288	289	257	337	2696	33462	304	176		
HPV 31	241	256	272	337	305	5168	161	289	240	208	209	321	256	289	321	353	337	288	305	401	241	337	241	289	305	289	273	305	305	273	3001	35901	257	113		
HPV 33	112	96	128	96	80	64	9245	113	96	48	161	96	0	256	96	96	48	128	177	80	81	49	80	96	144	128	64	112	97	32	12036	59597	16	80		
HPV 33	48	113	113	144	128	96	8940	16	113	112	128	112	16	80	97	160	128	129	144	96	64	128	129	145	48	96	96	128	161	113	12679	59597	80	161		
HPV 35	0	64	64	112	0	16	129	7142	0	96	48	0	161	80	32	33	144	80	64	48	80	32	96	16	128	64	80	304	16	6355	40395	64	48			
HPV 35	0	0	0	0	64	0	0	6885	0	48	0	0	0	0	0	0	0	0	0	32	0	16	0	33	0	0	289	0	6018	36704	0	0				
HPV 39	0	128	81	225	48	16	321	0	13819	32	16	80	0	65	32	161	144	0	112	176	80	240	80	208	97	96	224	32	112	112	6339	33061	0	16		
HPV 39	80	128	113	257	144	33	176	129	13834	113	64	145	32	161	65	145	112	16	144	193	113	128	96	192	128	161	192	48	144	128	6307	30526	48	16		
HPV 40	160	177	129	193	96	97	161	65	145	11057	193	144	64	81	80	225	160	65	144	112	129	144	160	160	128	177	176	128	144	112	9004	59581	16	16		
HPV 40	224	129	113	241	128	113	144	193	225	11651	64	112	48	16	113	193	193	129	144	144	161	144	128	224	161	257	160	48	80	144	9132	59485	0	64		
HPV 42	225	97	16	48	289	97	80	97	97	32	4654	80	48	64	145	0	129	96	0	64	80	144	80	112	64	0	80	144	97	145	145	2376	23014	128	64	
HPV 42	0	240	81	97	48	0	112	113	81	16	3194	0	112	0	113	48	65	64	417	80	48	128	64	128	64	48	64	81	48	96	2359	26305	113	112		
HPV 43	144	48	145	81	80	65	80	48	81	65	48	81	65	48	113	0	128	0	96	96	96	160	112	32	65	113	161	144	0	129	5810	49777	96	0		
HPV 43	128	49	49	80	0	64	64	129	16	32	5232	96	48	81	401	32	80	0	48	96	96	144	17	145	96	0	418	161	5970	49046	80	32				
HPV 44	96	112	145	161	112	33	96	0	65	16	129	96	3354	48	81	113	16	129	112	16	129	16	112	192	97	48	96	97	129	3515	27556	177	0			
HPV 44	16	145	97	81	128	129	0	80	32	192	81	80	2873	128	32	80	144	97	48	0	80	80	64	0	177	145	97	161	96	96	3146	22485	96	16		
HPV 45	144	145	81	17	16	16	48	16	16	81	0	80	97	4975	16	64	96	49	80	48	64	80	113	145	241	48	80	112	16	48	4413	24924	0	80		
HPV 45	49	16	48	32	0	0	0	32	48	32	64	3755	16	65	48	0	65	64	32	32	32	33	64	64	48	0	32	3434	22099	64	16					
HPV 51	0	48	64	32	32	16	0	16	0	48	32	80	16	1412	65	48	128	32	65	64	96	128	48	48	16	112	32	64	96	1220	10416	48	48			
HPV 51	80	0	64	16	97	0	209	48	0	48	32	64	64	0	1075	96	64	32	160	49	16	112	48	16	64	0	80	16	112	1204	11940	16	96			
HPV 52	193	96	96	160	176	0	128	209	129	80	96	129	145	0	97	9485	96	177	96	48	64	129	129	177	96	97	145	128	192	161	4976	41085	112	96		
HPV 52	129	176	64	112	32	49	0	64	17	32	144	17	65	16	33	8731	192	209	0	129	129	0	97	129	144	209	161	224	176	96	4494	38213	193	144		
HPV 53	65	0	0	0	321	321	0	16	0	32	0	0	0	0	0	5328	32	0	112	64	0	112	64	0	0	0	0	0	0	0	7961	38421	48	32		
HPV 53	32	16	0	48	96	0	16	0	33	16	0	80	0	80	16	5505	113	0	65	112	48	64	0	112	96	48	80	0	64	7768	34264	48	0			
HPV 54	144	0	113	0	112	97	64	49	129	97	16	96	112	65	32	96	32	10415	128	81	48	32	112	80	112	112	48	48	128	8891	59529	0	16			
HPV 54	16	97	16	81	32	64	64	16	112	0	112	16	0	321	32	113	9533	193	16	48	16	16	96	97	49	32	32	16	128	9646	59533</					

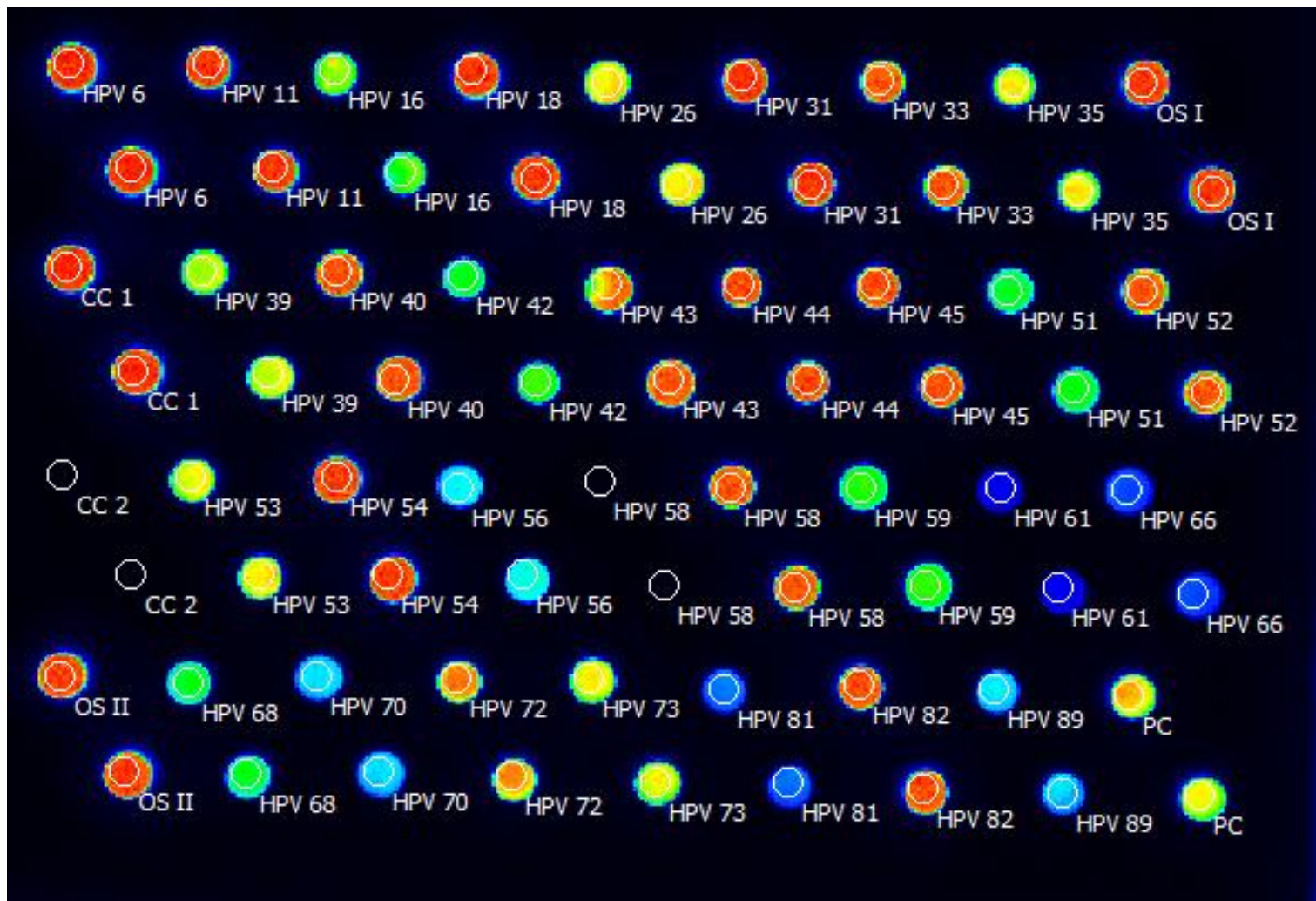
After various optimization: High sensitivity and specificity for 30 HPV



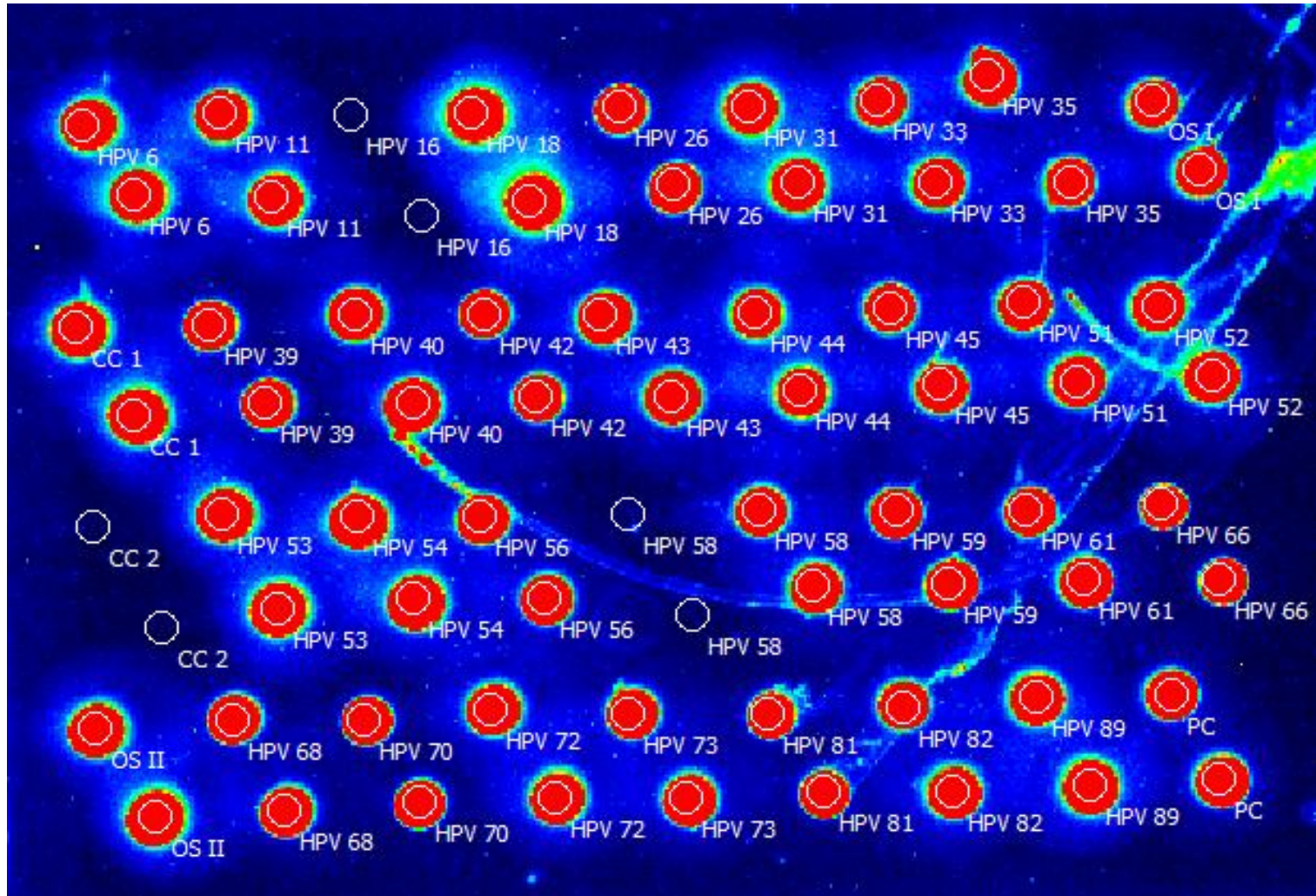
EUROArray HPV - High sensitivity and specificity for 30 HPV types



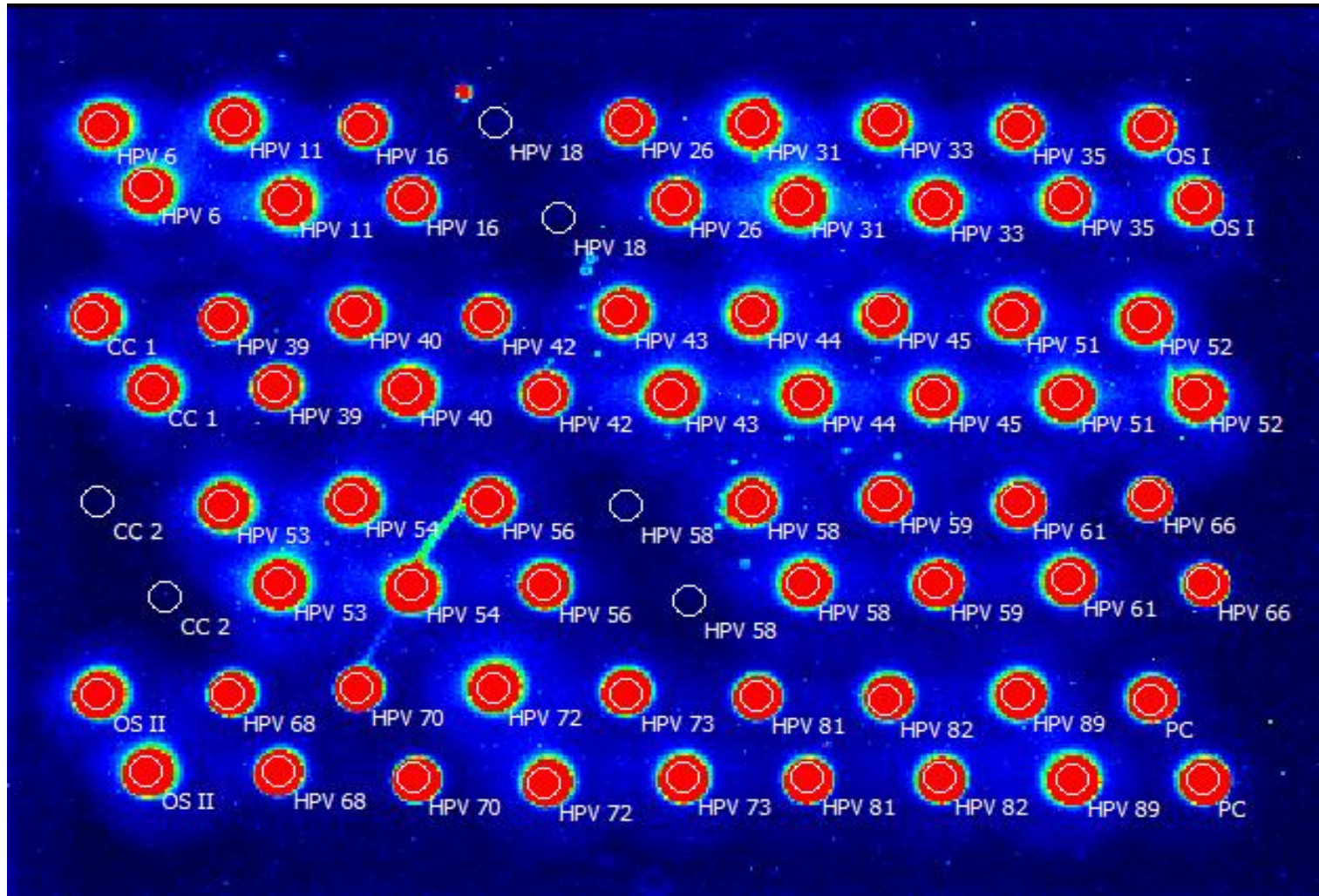
Detection and Typing of all 30 anogenital HPV at the LoD in one reaction



High specificity - EUROArray with all HPV but without HPV 16



High specificity EUROArray HPV PDM without HPV 18



Multiplex analysis make a detection and typing of HPV in one step possible

- Detection of all anogenital high-risk HPV
 - Multiple HPV- infections are common
(Menton et al., 2009; Insinga et al., 2008)
- Detection of all anogenital low-risk HPV
 - Differentiation between high-risk / low-risk induced CIN II
 - low-risk HPV may act as additional risk-factor for the development of cervical cancer



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Cervical cytology and multiple type HPV infection: A study of 8182 women ages 31–65[☆]

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HIGHLIGHTS

- Women over the age of 30 with multiple type HPV infections are more likely to have abnormal cytology.
- Women with multiple type HPV infections including HPV 16 had the highest OR associated with HSIL cytology.
- Continued study necessary to identify the impact of multiple type HPV infections on abnormal cytology



Reasons for HPV-subtyping

- Detection and typing in one step
 - Different HPV-subtypes come along with different risks
 - It is possible to discriminate between new and persistent infections
 - HPV-typing can be used as progression marker
 - The risk to develop cervical cancer is higher with multiple Infections, only with a typing test multiple infections are visible



Some questions only can be answered with multiplex parameter platforms

Detection of E6-E7 genes

- Not very conservative genes
- Detection of the oncogenes itself not of the capsid genes
- One primer-system for each HPV
- One specific probe for each HPV
- Detection also if the viral DNA is integrated in the host genom
 - A requirement for the malign transformation of the cell is the integration of the HPV-DNA into the human host- genom
(Hopman et al., 2004; Durst et al., 1985; Cullen et al., 1991; Hopman et al., 2006)



EUROArray Workflow

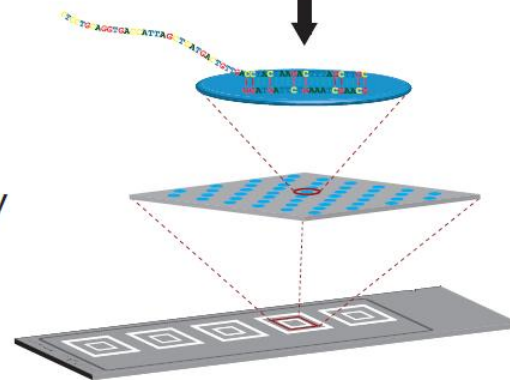
DNA sample



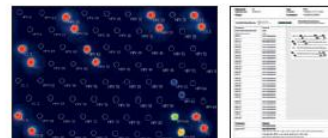
PCR (polymerase chain reaction)



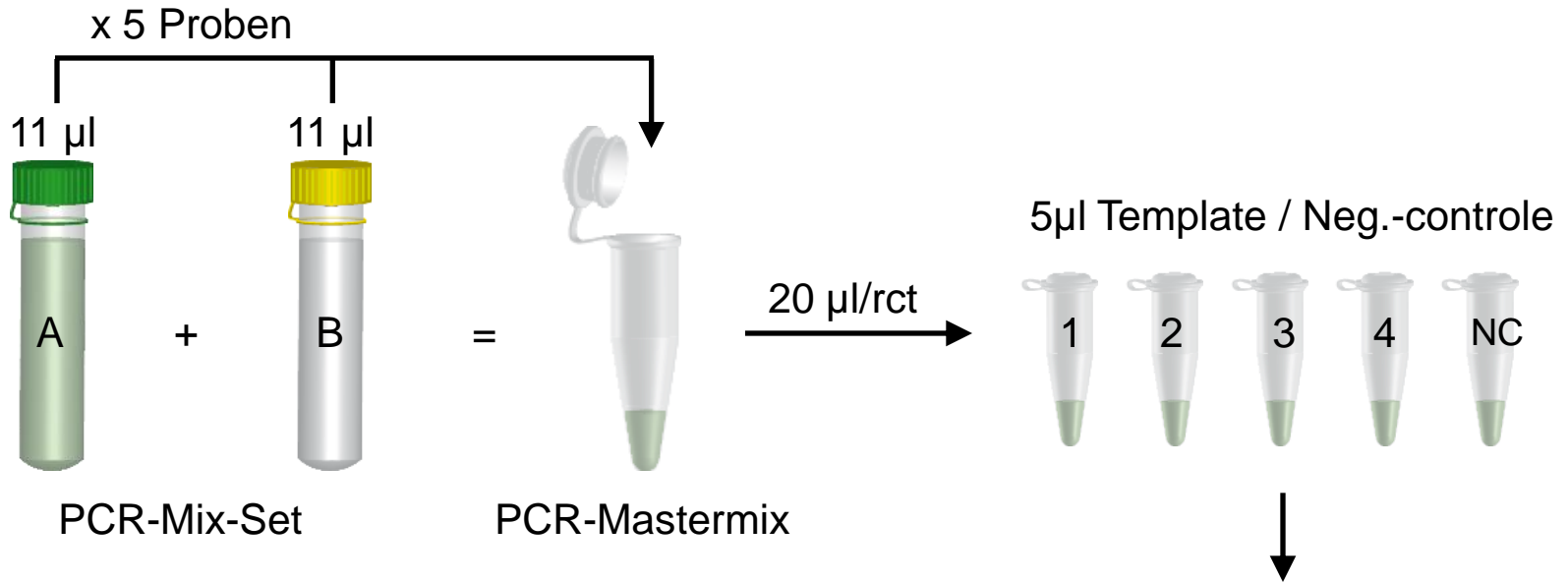
DNA microarray hybridisation



Fully automated evaluation



EUROArray system - Polymerase chain reaction



- Ready-to-use PCR components
- Very limited number of pipetting steps
- **Simple, fast, robust**



The PCR products are labeled during the reaction



Microarray hybridization

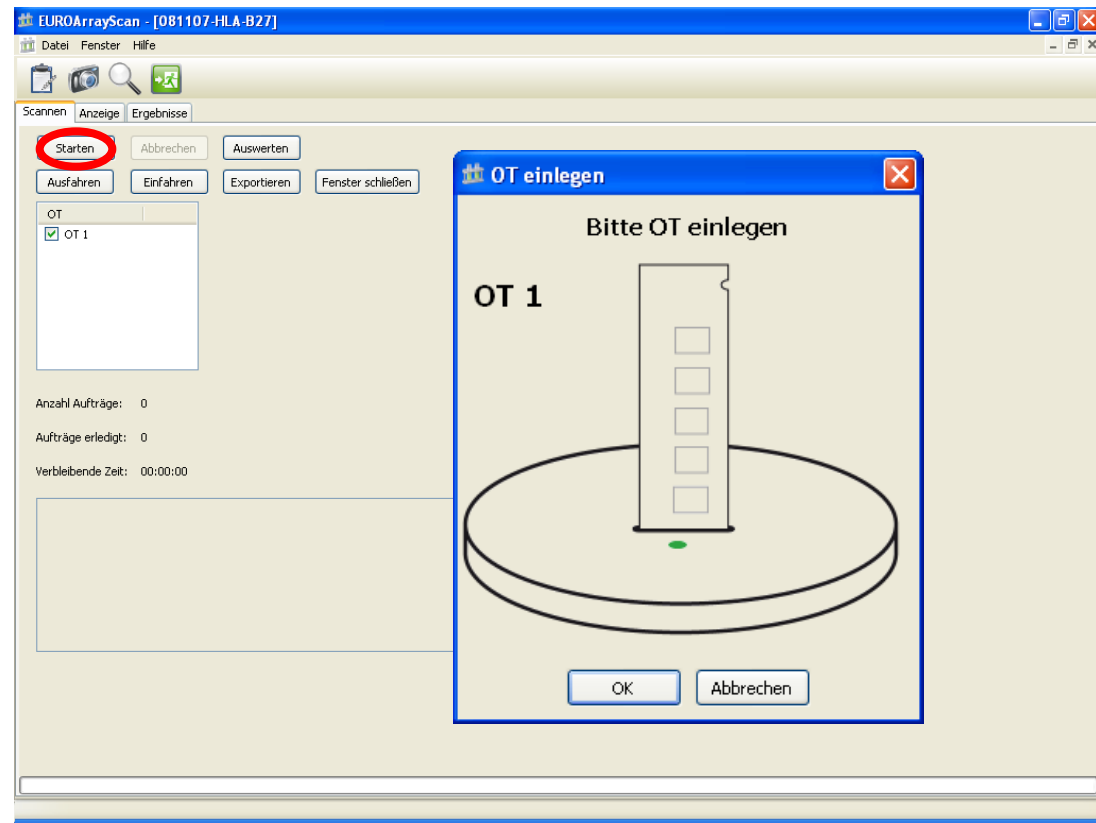
- Reproducible, simple handling



Scanning and evaluation

Fully automated standardized evaluation, interpretation and archiving of results

- Opening of the protocol and start
- Insertion of the μ Array slide

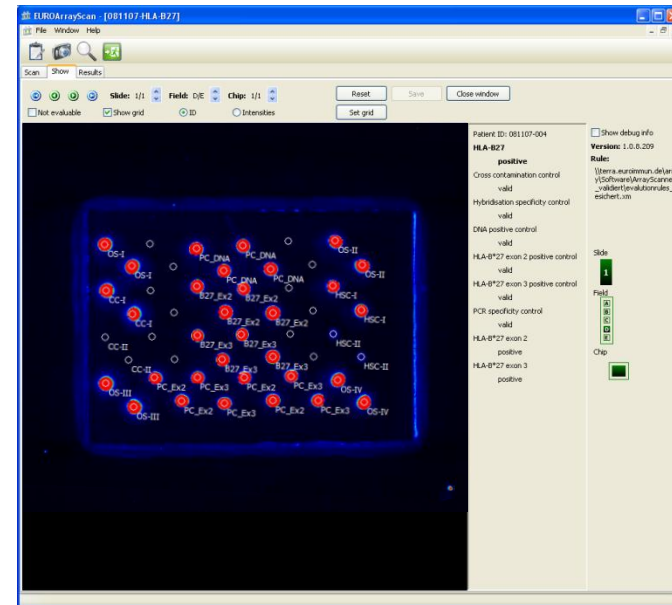


EUROArray system - Scanning and evaluation

Microarray Scanner



Software





- Complete process from receipt of samples to issuing of results is IVD validated and CE labelled (DNA extraction, test reagents, microarray scanner, software)



Patienten ID : Thinprep 23 ohne waschschritte
 Ergebnis vom : 22.10.2013
 Druckdatum : 15.01.2014 15:01:03

Test : HPV
 Protokoll : 131021JG_HP

EUROIMMUN <small>Medizinische Labordiagnostika AG</small>			Automatische Auswertung mit der EUROArrayScan-Software
		OT 1 Field C Chip 1	
Teilergebnis	Ergebnis		
Kreuz-Kontaminationskontrolle	valide		
DNA Positivkontrolle	valide		
HPV 6**	nicht nachgewiesen		
HPV 11**	nicht nachgewiesen		
HPV 16*	nicht nachgewiesen		
HPV 18*	nicht nachgewiesen		
HPV 26*	nicht nachgewiesen		
HPV 31*	nicht nachgewiesen		
HPV 33*	nicht nachgewiesen		
HPV 35*	nicht nachgewiesen		
HPV 39*	nicht nachgewiesen		
HPV 40**	nicht nachgewiesen		
HPV 42**	nicht nachgewiesen		
HPV 43**	nicht nachgewiesen		
HPV 44**	nicht nachgewiesen		
HPV 45*	nicht nachgewiesen		
HPV 51*	nicht nachgewiesen		
HPV 52*	nicht nachgewiesen		
HPV 53*	nicht nachgewiesen		
HPV 54**	nicht nachgewiesen		
HPV 56*	nicht nachgewiesen		
HPV 58*	NACHGEWIESEN		
HPV 59*	nicht nachgewiesen		
HPV 61**	nicht nachgewiesen		
HPV 66*	nicht nachgewiesen		
HPV 68*	nicht nachgewiesen		
HPV 70**	nicht nachgewiesen		
HPV 72*	nicht nachgewiesen		
HPV 73*	nicht nachgewiesen		
HPV 81**	nicht nachgewiesen		
HPV 82*	nicht nachgewiesen		
HPV 89**	nicht nachgewiesen		
Testergebnis	Ergebnis		
high-risk HPV*	NACHGEWIESEN		
low-risk HPV**	nicht nachgewiesen		
		*high-risk HPV: HPV 16, 18, 26, 31, 33, 35, 39, 45, 51, 52, 53, 56, 58, 59, 66, 68,73,82. **low-risk HPV: HPV 6, 11, 40, 42, 43, 44, 54, 61, 70, 72, 81, 89. Nach N Engl J Med 348:518-527 and Lancet Oncol 6(4):204.	

Unterschrift : _____





HPV microarray (IVD) – complete typing of human papilloma viruses



- Detection and typing of all 30 relevant anogenital HPV subtypes in one reaction
- Direct detection of the viral oncogenes E6/E7 provides highest possible sensitivity
- Significant results even in very early stages of the infection
- Distinction between high-risk and low-risk types of HPV
- Reliable identification of multiple infections

EUROArray HPV

(order no. MN 2540-####)



For detailed information about available test kit formats see our product catalogue or visit www.euroimmun.com.

- Well-established EUROArray technology
- Simple test performance – no in-depth molecular biology knowledge required!
- Ready-to-use PCR components, integrated controls
- Fully automated and standardised evaluation, interpretation and archiving of results (EUROArrayScan system)



Molecular diagnostics for HPV screening: sensitive, fast, reliable

- Alternative: molecular diagnosis
 - Sensitive
 - Fast
 - Typing of HPV subtypes is possible
 - Objective
 - Detection of the pathogen in a very early stage of the infection possible
 - Extremely sensitive
 - Sub-typing is possible
- A combination of PAP-Testing and HPV-Testing as cervical cancer prevention is recommended by the FDA and national and international associations of gynecologists

